Exact Stochastic Simulation of Coupled Chemical Reactions

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There are two formalisms for mathematically describing the time behavior of a spatially homogeneous chemical system: The deterministic approach regards the time evolution as a continuous, wholly predictable process which is governed by a set of coupled, ordinary differential equations (the "reaction-rate equations"); the stochastic approach regards the time evolution as a kind of random-walk process which is governed by a single differential-difference equation (the "master equation"). Fairly simple kinetic theory arguments show that the stochastic formulation of chemical kinetics has a firmer physical basis than the deterministic formulation, but unfortunately the stochastic master equation is often mathematically intractable. There is, however, a way to make exact numerical calculations within the framework of the stochastic formulation without having to deal with the master equation directly. It is a relatively simple digital computer algorithm which uses a rigorously derived Monte Carlo procedure to numerically simulate the time evolution of the given chemical system. Like the master equation, this "stochastic simulation algorithm" correctly accounts for the inherent fluctuations and correlations that are necessarily ignored in the deterministic formulation. In addition, unlike most procedures for numerically solving the deterministic reaction-rate equations, this algorithm never approximates infinitesimal time increments \( dt \) by finite time steps \( \Delta t \). The feasibility and utility of the simulation algorithm are demonstrated by applying it to several well-known model chemical systems, including the Lotka model, the Brusselator, and the Oregonator.

I. Introduction

In this paper we shall be concerned with the following general problem: If a fixed volume \( V \) contains a spatially uniform mixture of \( N \) chemical species which can interreact through \( M \) specified chemical reaction channels, then given the numbers of molecules of each species present at some initial time, what will these molecular population levels be at any later time?

The traditional way of treating this problem begins by translating it into the mathematical language of ordinary differential equations. More specifically, if we assume that the number of molecules of the \( i \)th species in \( V \) at time \( t \) can be represented by a continuous, single-valued function \( X_i(t) \) \( (i = 1 \ldots N) \) and if we further assume that each of the \( M \) chemical reactions can be regarded as a continuous rate process, then we can easily construct a set of coupled, first-order, ordinary differential equations of the form

\[
\frac{dX_1}{dt} = f_1(X_1, \ldots, X_N)
\]

\[
\frac{dX_2}{dt} = f_2(X_1, \ldots, X_N)
\]

\[
\frac{dX_N}{dt} = f_N(X_1, \ldots, X_N)
\]

(1)

The specific forms of the functions \( f_i \) on the right (which are usually nonlinear in the \( X_i \)’s) are determined by the structures and rate constants of the \( M \) chemical reaction channels. These equations are called the "reaction-rate equations"; solving them for the functions \( X_1(t), \ldots, X_N(t), \) subject to the prescribed initial conditions, is tantamount to solving the time-evolution problem posed earlier. Analytical solutions to the reaction-rate equations can be found only for rather simple systems, so it is usually necessary to solve these equations numerically on a computer. As is evident from many of the papers in this symposium series, the art of reliably solving reaction-rate equations on a computer has been developed to an impressive and sophisticated level.

Although the great importance and usefulness of the differential reaction-rate equations approach to chemical kinetics cannot be denied, we should not lose sight of the fact that the physical basis for this approach leaves something to be desired. This approach evidently assumes that the time evolution of a chemically reacting system is both continuous and deterministic. However, the time evolution of a chemically reacting system is not a continuous process, because molecular population levels obviously can change only by discrete integer amounts. Moreover, the time evolution is not a deterministic process either. For, even if we put aside quantum considerations and regard the molecular motions to be governed by the equations of classical mechanics, it is impossible even in principle to predict the exact molecular population levels at some future time unless we take account of the precise positions and velocities of all the molecules in the system. In other words, although the temporal behavior of a chemically reacting system of classical molecules is a deterministic process in the full position-momentum phase space of the system, it is not a deterministic process in the \( N \)-dimensional subspace of the species population numbers, as (1) implies.

In many cases of course the time evolution of a chemically reacting system can, to a very acceptable degree of accuracy, be treated as a continuous, deterministic process. However this should not always be taken for granted, especially now that the attention of chemical kineticists is increasingly being drawn to the study of ecological systems, microscopic biological systems, and nonlinear systems driven to conditions of chemical instability. In some cases like these, the inability of the reaction-rate equations to describe the fluctuations in the molecular population levels can be a serious shortcoming. Moreover, contrary to widespread belief, it is not even guaranteed that the reaction-rate equations will provide a sufficiently accurate account of the average molecular population levels; for, except for very simple linear systems, the average molecular population levels will not exactly satisfy any closed system of equations such as (1).
Many chemical kinetists would probably argue that these considerations are rather academic, because despite whatever shortcomings the differential reaction-rate equations may have, they seem to provide the only practical means for numerically analyzing the time behavior of nontrivial chemical systems. However, in a recent paper by this writer, a new computational method was developed from premises which take explicit account of the fact that the time evolution of a spatially homogeneous chemical system is a discrete, stochastic process instead of a continuous, deterministic process. This new computational method, which we shall refer to as the “stochastic simulation algorithm”, now offers an alternative to the differential reaction-rate equations—an alternative that is free of the difficulties mentioned above.

In sections II and III of this paper, we shall describe the stochastic simulation algorithm and give a brief synopsis of the arguments used in ref 1 to derive it. Then, in section IV, we shall exhibit for the first time some actual numerical results that have been obtained in applying the algorithm to some well-known model chemical systems. Our purpose in section IV will be not so much to shed new light on the models treated as to illustrate in a convincing way the feasibility and utility of our new computational method.

II. Physical Basis of the Stochastic Formulation of Chemical Kinetics

In general, a chemical reaction occurs whenever two or more molecules of appropriate kinds collide in an appropriate way. The “stochastic formulation” of chemical kinetics is simply a consequence of taking seriously the fact that collisions in a system of molecules in thermal equilibrium occur in an essentially random manner. In this section we want to sketch briefly how this intuitive notion can be expressed in a more precise, quantitative way; a more detailed treatment may be found in ref 1.

IIA. Molecular Collisions. Consider a system composed of a mixture of two gas-phase molecular species, S1 and S2, in thermal (but not necessarily chemical) equilibrium inside some volume V. For simplicity, let us assume that the S1 and S2 molecules are hard spheres of radii r1 and r2, respectively. Then a 1-2 collision will occur whenever the center-to-center distance between an S1 molecule and an S2 molecule decreases to r12 = r1 + r2. Let us try to calculate the rate at which such collisions are occurring in V. Following traditional textbook derivations of the molecular collision rate, we begin by picking an arbitrary 1-2 molecular pair, and denoting by v12 the speed of molecule 1 relative to molecule 2. We then observe that, in the next small time interval δt, molecule 1 will sweep out relative to molecule 2 a “collision volume” δVcoll = πr122v12δt (see Figure 1), in the sense that if the center of molecule 2 happens to lie inside δVcoll at time t, then the two molecules will collide in the time interval (t, t + δt).

The usual procedure now would be to estimate the number of S2 molecules whose centers lie inside δVcoll, divide that number by δt, and then take the limit δt → 0 to obtain the rate at which the S1 molecule is colliding with S2 molecules. However, this procedure suffers from an embarrassing difficulty: As δVcoll → 0, the number of S2 molecules whose centers lie inside δVcoll will be either 0 or 1, with the latter possibility becoming more and more likely as the limiting process proceeds. Strictly speaking, then, it is physically meaningless to talk about “the number of molecules whose centers lie inside δVcoll” in the required limit of vanishingly small δt.

Instead of following the usual practice of overriding this difficulty with nonrigorous “averaging” arguments, let us redirect our inquiry in the following way: Since the system is in thermal equilibrium, the molecules will at all times be distributed randomly and uniformly throughout the containing volume V. Therefore, the probability that the center of an arbitrary S2 molecule will be found inside δVcoll at time t will be given by the simple ratio δVcoll/V; moreover, this is true even in the limit of vanishingly small δVcoll. If we now average this ratio over the velocity distributions of the S1 and S2 molecules, we may conclude...
speed will be equal to Boltzmann's constant, \( k_B \). For Maxwellian velocity distributions the average relative velocity \( v_{12} \) that a particular 1-2 molecular pair will collide in the next vanishingly small time interval \( \delta t \) is given by:

\[
\delta V_{	ext{coll}} / V = V^{-1} \pi r_{12}^2 v_{12} \delta t
\]

where \( V \) is the volume, \( r_{12} \) is the distance between the centers of mass of the colliding molecules, \( v_{12} \) is their relative velocity, and \( \delta t \) is the infinitesimal time interval.

For Maxwellian velocity distributions the average relative speed \( v_{12} \) will be equal to \((8kT/(\pi m_{12}))^{1/2}\), where \( k \) is Boltzmann's constant, \( T \) the absolute temperature, and \( m_{12} \) the reduced mass \( m_1 m_2/(m_1 + m_2) \). In any case, if we are given that at time \( t \) there are in \( V \) \( X_1 \) of the \( S_1 \) molecules and \( X_2 \) of the \( S_2 \) molecules, making a total of \( X_1 X_2 \) distinct 1-2 molecular pairs, then it follows from (2a) that:

\[
X_1 X_2 V^{-1} \pi r_{12}^2 v_{12} \, dt = \text{probability that a 1-2 collision will occur somewhere inside } V \text{ in the next infinitesimal time interval } (t, t + dt)
\]

We see then that, although we cannot rigorously calculate the number of 1-2 collisions occurring in \( V \) in any infinitesimal time interval, we can approximately calculate the probability of a 1-2 collision occurring in \( V \) in any infinitesimal time interval. This means that we really ought to characterize a system of thermally equilibrated molecules by a "collision probability per unit time"—namely, the coefficient of \( dt \) in (2b)—instead of by a "collision rate". This is why these collisions constitute a stochastic Markov process instead of a deterministic rate process.

### IIB. The Stochastic Reaction Constant \( c_1 \)

If one applies the foregoing arguments specifically to reactive collisions (i.e., collisions which result in chemical alterations of the colliding molecules), one finds in a similar way that chemical reactions are more properly characterized by a "reaction probability per unit time" instead of a "reaction rate". Thus, suppose the \( S_1 \) and \( S_2 \) molecules in \( V \) can undergo the reaction

\[
R_1: \quad S_1 + S_2 \rightarrow 2S_1
\]

Then in direct analogy with (2a), we may assert the existence of a constant \( c_1 \), which depends only on the physical properties of the two molecules and the temperature of the system, such that

\[
c_1 \, dt = \text{average probability that a particular 1-2 molecular pair will react according to } R_1 \text{ in the next infinitesimal time interval } (t, t + dt)
\]

This implies that, if at time \( t \) there are in \( V \) \( X_1 \) of the \( S_1 \) molecules and \( X_2 \) of the \( S_2 \) molecules, making a total of \( X_1 X_2 \) distinct 1-2 pairs, then:

\[
X_1 X_2 c_1 \, dt = \text{probability that an } R_1 \text{ reaction will occur somewhere inside } V \text{ in the next infinitesimal time interval } (t, t + dt)
\]

More generally, suppose the volume \( V \) contains a spatially homogeneous (or thermally equilibrated) mixture of \( X_1 \) molecules of chemical species \( S_i \) \( (i = 1, \ldots, N) \), and suppose further that these \( N \) species can interreact through \( M \) specified chemical reaction channels \( R_\mu \) \( (\mu = 1, \ldots, M) \). Then we may assert the existence of \( M \) constants \( c_\mu \) \( (\mu = 1, \ldots, M) \), which depend only on the physical properties of the molecules and the temperature of the system, such that:

\[
c_\mu \, dt = \text{average probability that a particular combination of } R_\mu \text{ reactant molecules will react accordingly in the next infinitesimal time interval } (t, t + dt)
\]

By "average" here we mean simply that, if we multiply \( c_\mu \, dt \) by the total number of distinct combinations of \( R_\mu \) reactant molecules in \( V \) at time \( t \), we will obtain the probability that an \( R_\mu \) reaction will occur somewhere inside \( V \) in the next infinitesimal time interval \( (t, t + dt) \).

Equation 4 may be regarded both as the definition of the stochastic reaction constant \( c_\mu \); and also as the fundamental hypothesis of the stochastic formulation of chemical kinetics. As is shown by more detailed arguments in ref 1, we may expect this hypothesis to be valid for any molecular system that is kept "well-mixed", either by...
direct stirring, or else by simply requiring that nonreactive molecular collisions occur much more frequently than reactive molecular collisions. In section III we shall take up the problem of how to use (4) to actually calculate the time evolution of such a system.

IIIC. Connection with the Deterministic Reaction-Rate Constant \( k_s \). Intuition suggests that the stochastic reaction constant \( c_s \) should be closely related to the more familiar reaction-rate constant \( k_s \), which forms the basis for the deterministic approach to chemical kinetics. For the particular reaction \( R_1 \) in (3a), it turns out that

\[
\frac{\Delta V}{\Delta t} = Vc_1(X_1 X_2)/(X_1)(X_2) \tag{5a}
\]

where the angular brackets denote an average over an ensemble of identical systems. However, in the deterministic formulation of chemical kinetics, no distinction is made between the average of a product and the product of the averages. Therefore, to the extent that the deterministic formulation is adequate, we have \( \langle X_1 X_2 \rangle = \langle X_1 \rangle \langle X_2 \rangle \), and (5a) simplifies to

\[
\frac{\Delta V}{\Delta t} = Vc_1 \tag{5b}
\]

The presence of the factor \( V \) is a consequence of the fact that the reaction-rate constant is normally used in differential equations that contain molecular concentrations (numbers of molecules per unit volume) rather than total numbers of molecules. If reaction \( R_1 \) had three reactant molecules instead of two, we would have \( V^2 \) instead of \( V \); if \( R_1 \) had only one reactant molecule (a simple isomerization), the factor \( V \) would be absent.

Aside from possible factors of \( V \), there is one other practical difference between \( c_s \) and \( k_s \) that can arise. To see this, consider the reverse of the reaction \( R_1 \) in (3a):

\[
R_1: \quad 2S \rightarrow S_1 + S_2 \tag{6a}
\]

Following (4), we would characterize this reaction by a constant \( c_s \) such that \( c_s \Delta t \) is the average probability that a particular pair of \( S \) molecules will react according to \( R_1 \) in the next \( \Delta t \). However, the number of distinct pairs of \( S \) molecules in \( V \) is not \( X_1 X_2 \) but \( X_1(X_1 - 1)/2! \). Hence, the probability that an \( R_2 \) reaction will occur somewhere inside \( V \) in the next time interval \( \Delta t \) is \( 1/2 X_1(X_1 - 1)c_s \Delta t \). This leads to the result that

\[
k_s = Vc_1\langle 1/2 X_1(X_1 - 1)/\langle X_1 \rangle \rangle = Vc_1/2 \tag{6b}
\]

in contrast to eq 5. In general, if \( R_2 \) has two identical reactant molecules then \( c_s \) will be larger than \( k_s \) by a factor of \( 2! = 2 \); if \( R_2 \) has three identical reactant molecules, this factor will be \( 3! = 6 \).

We see then that, from a practical numerical point of view, \( c_s \) and \( k_s \) will differ at most by only two simple constant factors. From a theoretical point of view, however, the difference between \( c_s \) and \( k_s \) is much more complicated, and relates ultimately to the conceptual differences that exist between the stochastic and deterministic approaches to chemical kinetics.

III. Calculating the Stochastic Time Evolution of a Chemically Reacting System

The temporal behavior of a spatially homogeneous mixture of \( N \) molecular species interreacting through \( M \) chemical reaction channels is governed solely by the fundamental hypothesis (4). However, it is not at all obvious just how we should go about using (4) to calculate this temporal behavior. In this section we shall outline two different ways of approaching this problem—the traditional “master equation” approach, and the new “stochastic simulation” approach. Although their strategies are quite different, it is important to keep in mind that these two approaches are nevertheless equivalent in the sense that both are rigorous consequences of the fundamental hypothesis (4).

III. The Master Equation Approach. The traditional method of calculating the stochastic time evolution of a chemically reacting system is to set up and solve a so-called “master equation” for the system. A good review of the master equation approach to chemical kinetics has been given by McQuarrie. Here we would merely like to summarize briefly the main features of the master equation formalism in order to provide a conceptual setting for our subsequent presentation of the stochastic simulation approach. It should be emphasized, though, that the master equation itself plays no role whatsoever in either the derivation or the implementation of the stochastic simulation algorithm.

The key element of the master equation formalism is the “grand probability function”

\[
P(X_1, X_2, \ldots, X_N; t) = \text{probability that there will be in } V \text{ at time } t \\
\text{molecules of species } S_1, \text{ and } X_2 \text{ molecules of species } S_2, \ldots, \text{ and } X_N \text{ molecules of species } S_N
\]

A knowledge of this function would evidently provide a fairly complete characterization of the “stochastic state” of the system at time \( t \). Thus, for example, the \( k \)th moment of \( P \) with respect to \( X_i \)

\[
X_i^{(k)}(t) = \sum_{X_i=0}^\infty \sum_{X_N=0}^\infty X_i^k P(X_1, \ldots, X_N; t)
\]

(8)

gives the “average (number)” of \( S_i \) molecules in \( V \) at time \( t \). By “average” here we mean an average taken over many repeated “runs” from time 0 to time \( t \) of the stochastic process defined by (4), each run having the same initial numbers of molecules. The number \( X_i \) of \( S_i \) molecules found at time \( t \) will vary from run to run, but the average of the \( k \)th power of these numbers will approach \( X_i^{(k)}(t) \) in the limit of infinitely many runs. Especially useful are the \( k = 1 \) and \( k = 2 \) moments; this is because \( X_i^{(1)}(t) \) and

\[
\Delta X_i(t) = \{ X_i^{(0)}(t) - [X_i^{(0)}(t)]^2 \}^{1/2}
\]

measure, respectively, the average number of \( S_i \) molecules
in $V$ at time $t$, and the magnitude of the root-mean-square fluctuations that occur about this average. In other words, we may "reasonably expect" to find between
\[ X^{(1)}(t) - \Delta(t) \quad \text{and} \quad X^{(1)}(t) + \Delta(t) \]
(10)
molecules of $S_i$ in $V$ in time $t$. The functions $X(t)$ appearing in the deterministic reaction-rate equations (1) usually approximate the first moments $X^{(1)}(t)$ fairly closely, but only rarely is there exact equality.

The master equation is simply the time-evolution equation for the function $P(X_1, ..., X_N; t)$. It may be derived from (4) by using the addition and multiplication laws of probability theory to write $P(X_1, ..., X_N; t + dt)$ as the sum of the probabilities of the $1 + M$ different ways in which the system can arrive at the state $(X_1, ..., X_N)$ at time $t + dt$.

$$P(X_1, ..., X_N; t + dt) = P(X_1, ..., X_N; t)[1 - \sum_{\mu=1}^{M} a_{\mu} dt] + \sum_{\mu=1}^{M} B_{\mu} dt$$
(11)

Here we have defined the quantities $a_{\mu}$ by

$$a_{\mu} dt = c_{\mu} dt \times \text{(number of distinct } R_\mu \text{ molecular combinations in the state } (X_1, ..., X_N))$$

= probability that an $R_\mu$ reaction will occur in $V$ in $(t, t + dt)$, given that the system is in the state $(X_1, ..., X_N)$ at time $t$ (12)

Thus, the first term in (10) is the probability that the system will be in the state $(X_1, ..., X_N)$ at time $t$, and then remains in that state (i.e., undergoes no reactions) in $(t, t + dt)$. The quantity $B_{\mu} dt$ gives the probability that the system is one $R_\mu$ reaction removed from the state $(X_1, ..., X_N)$ at time $t$, and then undergoes an $R_\mu$ reaction in $(t, t + dt)$. Thus, $B_{\mu}$ will be the product of $P$ evaluated at the appropriate one-removed state at $t$, times $c_{\mu}$ times the number of $R_\mu$ molecular reactant combinations available in that one-removed state. Without going into any further details, it is sufficient here to simply observe that (11) leads directly to the "master equation"

$$\frac{\partial}{\partial t}P(X_1, ..., X_N; t) = \sum_{\mu=1}^{M} [B_{\mu} - a_{\mu} P(X_1, ..., X_N; t)]$$
(13)

In any particular case, the master equation is fairly easy to write; however, solving it is quite another matter. The number of problems for which the master equation (13) can be solved analytically is even fewer than the number of problems for which the deterministic reaction-rate equations (1) can be solved analytically. In addition, unlike the reaction-rate equations, the master equation does not readily lend itself to numerical solution on a digital computer, owing to the number and nature of its independent variables. Attempts to use the master equation to construct tractable time-evolution equations for the moments, and in particular $X^{(1)}(t)$ and $\Delta(t)$, are also usually unsuccessful; for it turns out that, unless all the reactions $R_i$ are simple unimolecular reactions, the equations for the time derivative of any moment will always involve higher order moments, thus rendering the set of moment equations infinitely open ended.

In short, although the master equation is both exact and elegant, it is usually not very useful for making practical numerical calculations.

III. The Reaction Probability Density Function. Let us now put aside the master equation formalism, and consider instead how we might go about simulating the stochastic time evolution of a chemically reacting system. If we are given that the system is in the state $(X_1, ..., X_N)$ at time $t$, then essentially all we need in order to "move the system forward in time" are the answers to two questions: when will the next reaction occur, and what kind of reaction will it be? Because of the stochastic nature of the reactions, though, we may expect that these two questions will be answerable only in some "probabilistic" sense.

Prompted by these considerations, we introduce the function $P(\tau, \mu)$ defined by

$$P(\tau, \mu) d\tau = \text{probability that, given the state } (X_1, ..., X_N) \text{ at time } t, \text{ the next reaction in } V \text{ will occur in the infinitesimal time interval } (t + \tau, t + \tau + d\tau), \text{ and will be an } R_\mu \text{ reaction } (14)$$

We call $P(\tau, \mu)$ the "reaction probability density function", because in mathematical terminology it is a joint probability density function on the space of the continuous variable $\tau (0 \leq \tau < \infty)$ and the discrete variable $\mu (\mu = 1, 2, ..., M)$. Notice that the variables $\tau$ and $\mu$ are quantities whose respective values would give us answers to the two questions mentioned above. Our first step toward finding a legitimate method for assigning numerical values to $\tau$ and $\mu$ is to derive, from the fundamental hypothesis (4), an analytical expression for $P(\tau, \mu)$.

To this end, we begin by defining for each reaction $R_\mu$ a function $h_\mu$ according to

$$h_\mu = \text{number of distinct } R_\mu \text{ molecular reactant combinations available in the state } (X_1, ..., X_N) \text{ at time } t \quad (\mu = 1, ..., M)$$
(15)

Thus, if $R_\mu$ has the form $S_1 + S_2 \rightarrow$ anything, then we will have $h_\mu = X_1X_2$; if $R_\mu$ has the form $2S_1 \rightarrow$ anything, then we will have $h_\mu = \frac{X_1}{2}(X_1 - 1)$. In general, $h_\mu$ will be some combinatorial function of the variables $X_1, X_2, ..., X_N$. With $h_\mu$ so defined, then (4) implies that [cf. (12)]

$$a_{\mu} dt = h_\mu c_{\mu} dt = \text{probability that an } R_\mu \text{ reaction will occur in } V \text{ in } (t, t + dt), \text{ given that the system is in the state } (X_1, ..., X_N) \text{ at time } t \quad (\mu = 1, ..., M)$$
(16)

We now calculate the probability in (14) as the product of: $P(\tau)$, the probability that, given the state $(X_1, ..., X_N)$ at time $t$, no reaction will occur in the time interval $(t, t + \tau)$; times $a_{\mu} d\tau$, the subsequent probability that an $R_\mu$ reaction will occur in the time interval $(t + \tau, t + \tau + d\tau)$; $P(\tau, \mu) d\tau$. To find an expression for $P(\tau)$, we first note that $[1 - \Sigma a_{\mu} d\tau]$ is the probability that no reaction will occur in time $\tau$ from the state $(X_1, ..., X_N)$. Therefore

$$P(\tau) = \prod_{\mu=1}^{M} [1 - \Sigma a_{\mu} d\tau]$$
(17a)

from which it is readily deduced that

$$P(\tau) = \exp[-\Sigma a_{\mu} \tau]$$
(17c)

Inserting (17c) into (17a), we conclude that the reaction probability density function defined in (14) is given by

$$P(\tau, \mu) = \begin{cases} a_{\mu} \exp(-a_{\mu} \tau) & \text{if } 0 \leq \tau < - \text{ and } \mu = 1, ..., M \\ 0 & \text{otherwise} \end{cases}$$
(18)

where

$$a_{\mu} = h_\mu c_{\mu} \quad (\mu = 1, ..., M)$$
(19a)

and

$$a_{\mu} = \Sigma_{\mu=1}^{M} a_{\mu} = \Sigma_{\mu=1}^{M} h_\mu c_{\mu}$$
(19b)
The expression for \( P(\tau, \mu) \) in (18) is, like the master equation (13), a rigorous mathematical consequence of the fundamental hypothesis (4). Notice that \( P(\tau, \mu) \) depends on all the reaction constants (not just on \( c_i \)), and on the current numbers of molecules of all reactant species (not just on the \( R_i \) reactants).

III. The Stochastic Simulation Algorithm. At the beginning of section IIIB, we observed that essentially what is needed to simulate the time evolution of a chemically reacting system is some way of specifying when the next reaction will occur and what kind of reaction it will be. We can now frame this requirement in more precise, mathematical terms: What is needed is a method for drawing or "generating" a pair \((\tau, \mu)\) from the set of random pairs whose probability density function is \( P(\tau, \mu) \) in (18). It turns out that there is a simple, rigorous way of doing this on a digital computer, provided we have access to a reliable "unit-interval uniform random number generator".

A unit-interval uniform random number generator is a computer subprogram which, when called, calculates and returns a random (or more properly, a "pseudorandom") number \( r \) from the uniform distribution in the unit interval; that is, the a priori probability that any generated number \( r \) will fall inside any given subinterval \([a, b]\) of the unit interval \([0, 1]\) is equal to \( b - a \). Nowadays, virtually every large digital computer facility has one of these subprograms in its library file that is fast, easy to use, and sufficiently reliable. When called, the typical generator subprogram calculates the required pseudorandom number \( r \) by performing a few relatively simple operations on the previous pseudorandom number that was calculated. Thus, \( n \) successive calls will produce a "chain" of \( n \) pseudorandom numbers. The first number of the chain is determined by initializing the generator with a user-chosen starter number; different starter numbers will result in different but statistically equivalent chains.

Now, if our goal were to generate a random pair \((\tau, \mu)\) according to the probability density function

\[
P(\tau, \mu) = \begin{cases} 
1 & \text{if } 0 \leq \tau \leq 1 \text{ and } 0 \leq \mu \leq 1 \\
0 & \text{otherwise}
\end{cases} \tag{20a}
\]

then we could simply generate two random numbers \( r_1 \) and \( r_2 \) using a unit-interval uniform random number generator and take

\[
\tau = r_1 \quad \mu = r_2 \tag{20b}
\]

However, our objective here is to generate a random pair \((\tau, \mu)\) according to the probability density function in (18), not the probability density function in (20a). As it happens, there exists a mathematically rigorous procedure for taking two random numbers \( r_1 \) and \( r_2 \) from the unit-interval uniform distribution, and constructing from them a random pair \((\tau, \mu)\) from a set described by any specified pair probability density function. For the pair probability density function in (20a), this construction procedure turns out to be precisely (20b); for the pair probability density function \( P(\tau, \mu) \) in (18), the construction procedure turns out to be as follows:

With \( r_1 \) and \( r_2 \) two random numbers from the unit-interval uniform distribution, take

\[
\tau = \left(1/a_0\right) \ln \left(1/r_1\right) \tag{21a}
\]

and take \( \mu \) to be that integer for which

\[
\sum_{\mu=1}^{\mu} a_\mu < r_2 a_0 \leq \sum_{\mu=1}^{\mu+1} a_\mu \tag{21b}
\]

A rigorous proof of the fact that the pair \((\tau, \mu)\) constructed according eq 21 may be regarded as having been drawn from the set of random pairs whose probability density function is \( P(\tau, \mu) \) in (18) may be found in section VA of ref 1. Suffice it here to say that (21a) generates a random number \( \tau \) according to the probability density function \( P_1(\tau) = a_0 \exp(-a_0 \tau) \), while (21b) generates a random integer \( \mu \) according to the probability density function \( P_2(\mu) = a_\mu/a_0 \), and the stated result follows, roughly speaking, because \( P(\tau)P_2(\mu) = P(\tau, \mu) \).

The generating procedure (21) is easy to code in Fortran. In particular, (21b) may be implemented simply by cumulatively adding the successive values \( a_1, a_2, \ldots \) in a do-loop until their sum is observed to equal or exceed \( r_2 a_0 \), whereupon \( \mu \) is set equal to the index of the last \( a_\mu \) term added.

Our algorithm for simulating the stochastic time evolution of a chemically reacting system should now be rather obvious (see Figure 2):

**Step 0 (Initialization).** Input the desired values for the \( M \) reaction constants \( c_1, \ldots, c_M \) and the \( N \) initial molecular population numbers \( X_{10}, \ldots, X_{N0} \). Set the time variable \( t \) and the reaction counter \( n \) both to zero. Initialize the unit-interval uniform random number generator (URN).

**Step 1.** Calculate and store the \( M \) quantities \( a_1 = h_1 c_1, \ldots, a_M = h_M c_M \) for the current molecular population numbers, where \( h_i \) is that function of \( X_{10}, \ldots, X_{N0} \) defined in (15). Also calculate and store as \( a_0 \) the sum of the \( M \) \( a_\mu \) values.

**Step 2.** Generate two random numbers \( r_1 \) and \( r_2 \) using the unit-interval uniform random number generator, and calculate \( \tau \) and \( \mu \) according to (21a) and (21b).

**Step 3.** Using the \( \tau \) and \( \mu \) values obtained in step 2, increase \( t \) by \( \tau \), and adjust the molecular population levels to reflect the occurrence of one \( R_\mu \) reaction; e.g., if \( R_\mu \) is the reaction in (3a), then increase \( X_1 \) by 1 and decrease \( X_2 \) by 1. Then increase the reaction counter \( n \) by 1 and return to step 1.

In returning to step 1 from step 3, notice that it is necessary to recalculate only those quantities \( a_\mu \) corresponding to reactions \( R_\mu \) whose reactant population levels were just altered in step 3; also, \( a_0 \) may be recalculated simply by adding to \( a_0 \) the difference between each newly changed \( a_\mu \) value and its corresponding old value.

Of course, somewhere in the 1–2–3 loop one will want to provide for writing out or plotting the \( (X_1, \ldots, X_N, t) \) values at regular intervals of either \( t \) or \( n \). Also, one will want to make provisions for halting the calculations when either \( t \) or \( n \) reaches some predetermined value, or if \( a_0 \) should ever reach zero.

If it is desired to estimate the moment of the moments \( X_1(k)(t) \) of the grand probability function (see eq 8–10), then it will be necessary to make several simulation runs from time 0 to the chosen time \( t \), all identical with each other except for the initialization of the random number generator in step 0. Any moment \( X_1(k)(t) = \langle X_1^k \rangle \) may then be estimated directly as the average of the \( k \)th power of the numbers found for \( X_1 \) at time \( t \) in these runs. In a similar way, one can estimate various cross-correlation functions such as \( \langle X_1 X_2^l(\cdot) \rangle - \langle X_1(\cdot) \rangle \langle X_2(\cdot) \rangle \). The number of runs necessary to obtain adequate statistics in these estimates will vary with the situation.

IIIID. Remarks. After the simulation algorithm described above had been developed, several earlier simulation procedures were brought to the attention of the author. Two of these deserve to be mentioned here.

First is the method used by Nakashishi\(^{16}\) in 1972 to simulate the oscillating Lotka reactions. Like the computational method described above, Nakashishi’s method is expressly designed to numerically simulate the stochastic
process described by the spatially homogeneous master equation. However, Nakanishi's computational algorithm is neither as exact nor as efficient as ours. Essentially, it divides the time scale up into small bins $\Delta t$ and then decides on the basis of a random number which, if any, of the $M$ reaction types should be made to occur in each $\Delta t$. This approximate procedure becomes exact in the limit $\Delta t \to 0$, but unfortunately the efficiency of the procedure becomes nil in that same limit.

Much closer to our algorithm from an operational standpoint is the computational scheme proposed by Bunker et al. in 1974. In terms of our notation, Bunker et al. use the same $\mu$-selection process as (21b), but they replace the $\tau$-selection process in (21a) by simply $\tau = 1/\alpha_0$. Since $1/\alpha_0$ is precisely the mean of the $\tau$ values generated according to (21a), this substitution is a very reasonable approximation. Bunker et al. correctly characterize their computational procedure as a "hybrid method, intermediate between differential equation solution and Monte Carlo". However, they derive their method in a rather heuristic way within the context of the deterministic formalism instead of the stochastic formalism, and it is not at all clear from their development in precisely what respects their method should be regarded as approximate. From the standpoint of our exact computational procedure, though, the method of Bunker et al. appears to be somewhat better than its inventors may have supposed.

The stochastic simulation algorithm described in this paper has both its advantages and its limitations. Although these can be more readily appreciated in the context of the specific applications presented in the next section, we shall now summarize briefly the principle strengths and weaknesses of this computational approach.

On the plus side, the method has the following advantages:

1. The stochastic simulation algorithm is exact, in the sense that it takes full account of the fluctuations and correlations implied by the fundamental hypothesis (4). This is a consequence of the fact that, like the spatially homogeneous master equation, the stochastic simulation algorithm has been derived from (4) in a mathematically rigorous way.

2. Unlike standard numerical methods for numerically solving systems of differential equations, the stochastic simulation algorithm never approximates infinitesimal time increments $\Delta t$ by finite time steps $\Delta t$. This will be especially advantageous when dealing with systems in which the population levels can change suddenly and sharply with time.

3. As is evident from Figure 2, the stochastic simulation algorithm is very easy to code in Fortran for any specified set of chemical reactions, no matter how complicated or highly coupled these reactions may be.

4. A stochastic simulation computer program will normally require very little in the way of computer memory space. For $N$ molecular species and $M$ reaction channels, the principle variables to be stored are the $NX$ values, the $MC$ values, and the $M + 1$ $a_i$ values—a total of only $N + 2M + 1$ in all. Notice in particular that, although the amount of computer memory required does depend on the number of species $N$, it does not depend on the number of molecules of these various species.

5. In terms of the "statistical ensemble of systems" envisioned in the master equation approach, the stochastic simulation algorithm essentially provides us with information on the time behavior of individual ensemble members, such as one would observe in an ideal laboratory experiment. This complements quite nicely the information on the time behavior of ensemble averages which the master equation approach provides through the various moments of the grand probability function $P$. (25)

6. When ensemble averages (e.g., means, variances, correlations) cannot be readily calculated from the master equation, the stochastic simulation approach offers a universally straightforward (though possibly time-consuming) way of numerically estimating them.

On the minus side, the stochastic simulation algorithm has the following limitations:

1. A stochastic simulation computer program often requires lots of computer time to execute. From a cost standpoint, this fact is somewhat offset by the fact that such programs typically occupy only a small amount of computer memory. Nevertheless, the stochastic simulation algorithm places a high premium on the speed of the computer's central arithmetical unit.

2. This computer time limitation essentially limits the total number of molecular reactions that can be simulated; indeed, the execution time for a given set of reactions will be roughly proportional to that number. This in turn implies limits on the total numbers of molecules involved and/or the total system time that can be simulated. In other words, the stochastic simulation algorithm is restricted to distinctly finite chemical systems. However, in view of the fact that chemical reactions are usually rather "local" affairs, even for systems which as a whole contain enormous numbers of molecules, it is conceivable that this limitation might in some cases be more of an advantage than a disadvantage.

3. The stochastic simulation algorithm requires a "reliable" unit-interval uniform random number generator; however, a specific criterion for "reliability" is hard to come by. Many of the generators available today are quite good, and are probably quite adequate for our stochastic simulation algorithm, but the possibility of trouble from this source should never be ignored.

4. The stochastic simulation approach requires making several runs to estimate ensemble averages. Depending on the dynamical quantity whose ensemble average is desired, and also the statistical accuracy required of its estimate, the number of runs needed for averaging may run afoul of computer time/cost limitations.

IV. Illustrative Applications of the Algorithm

We shall now present some numerical results that have been obtained in applying the stochastic simulation algorithm to several spatially homogeneous model chemical systems. Although our results may provide a few new insights into some of the systems treated; it should be kept in mind that the systems themselves are not the primary focus of our discussion here; rather, our principle objective is to demonstrate that the stochastic simulation algorithm is a feasible and potentially useful method for numerically calculating the temporal behavior of coupled chemical reactions. We shall begin by considering some fairly simple systems, and then proceed to three successively more complicated systems, namely, the Lotka reactions, the Brusselator, and the Oregonator.

IVA. Some Simple Systems. Perhaps the simplest nontrivial "set of coupled chemical reactions" that one can write is the irreversible isomerization (or radioactive decay) reaction

$$X \rightarrow Z$$

(22)

This is a pedagogically attractive first problem to tackle because it can be solved analytically, without approximations, in both the deterministic and the stochastic formulations. In the deterministic formulation we have
the reaction-rate equation [cf. (1)]

\[ \frac{dX}{dt} = -cX \]  

(23a)

whose solution, given that \( X = X_0 \) at \( t = 0 \), is

\[ X(t) = X_0 e^{-ct} \]  

(23b)

In the stochastic formulation, we may easily derive the master equation [cf. (13)]

\[ \frac{d}{dt} P(X; t) = c \delta_{x,x_0}(X + 1)P(X + 1; t) - XP(X; t) \]  

(24a)

where the “Kronecker epsilon” \( \delta_{i,j} \) is by definition zero if \( i = j \) and unity if \( i \neq j \). This equation can be solved for the initial condition \( P(X; 0) = \delta_{x,x_0} \), by dealing with it successively for \( X = X_0, X = X_0 - 1, X = X_0 - 2, \) etc. In this way we find the solution

\[ P(X; t) = \frac{X_0!}{X!(X_0 - X)!} e^{-ct} \left( 1 - e^{-ct} \right)^{X_0 - X} \]  

(24b)

\( X = 0, 1, \ldots, X_0 \)

This is in the standard form of a binomial or Bernoulli probability function, and it is not difficult to calculate its mean and rms deviation directly from the definitions (8) and (9). They turn out to be

\[ X(1)(t) = X_0 e^{-ct} \]  

(24c)

and

\[ \Delta(t) = [X_0 e^{-ct} (1 - e^{-ct})]^{1/2} \]  

(24d)

The fact that the stochastic mean \( X(1)(t) \) in (24c) is exactly equal to the deterministic function \( X(t) \) in (22b) will rarely be true for more complicated chemical systems.

To apply our stochastic simulation algorithm to reaction 22, we put \( M = N = 1, c_1 = c, X_1 = X \), and \( h_1 = X \) in Figure 2 (and we also note that the \( r_i \) calculations in step 2 can be omitted). In Figure 3 we show, for

\[ c = 0.5, X_0 = 1000 \]  

(25)

how the results found in a typical stochastic simulation run of reaction 22 compare with the predictions of the master equation. The two dashed curves in Figure 3 are \( X(1)(t) \pm \Delta(t) \) (the one-standard deviation envelope), and the two solid curves are \( X(1)(t) \pm 2 \Delta(t) \) (the two-standard deviation envelope). The dots, of course, represent the trajectory of the stochastic simulation run. A dot has been plotted at the \((t, X)\) value found immediately after the occurrence of each molecular reaction. No attempt has been made to smooth these dots with a continuous connecting curve, so they essentially represent the “raw output” of the simulation run. The horizontal spacing between the dots gives a direct measure of the time separation between successive reactions, and as we should expect, the average time separation between successive isomerizations (decays) increases as the number of \( X \) molecules decreases. Because there were 1000 \( X \) molecules at \( t = 0 \), a total of 1000 reactions are simulated here, requiring exactly 1000 passes through the 1-2-3 loop of Figure 2. In this particular simulation run, all but 6 of the \( X \) molecules had isomerized (decayed) by \( t = 10 \); the last \( X \) molecule did not disappear until \( t = 15.6 \). The unit of time will depend upon the units assigned to the reaction constant \( c \), and is irrelevant to our purposes here.

The procedure used to obtain this plot was as follows: First, the computational steps outlined in Figure 2 were Fortran coded into a simulation program for reaction 22; provisions were made in this program so that the current \((t, X)\) value would be written out on a mass-storage file after every reaction, and also on paper after every five reactions. This simulation program was then executed on a Univac 1110 computer, and the resulting paper output was examined for the purpose of determining how the data should be plotted. Then, a plotting program was written to read off the data from the mass-storage file and construct the plot. The plotting program consisted primarily of Fortran calls to appropriate subroutines from the standard Displa (Display Integrated Software System and Plotting Language) library. Execution of the plotting program on the Univac 1110 computer caused the plot to be constructed on a device called an FR-80, which essentially is a cathode ray tube whose screen image is automatically recorded on photographic film; the resulting

Figure 8. Results of a \( 1 \times 10^9 \) reaction stochastic simulation run of the Lotka reactions (38), with \( c_1X = 10, c_2 = 0.01, \) and \( c_3 = 10 \), corresponding to the deterministic steady-state \( Y_1 = Y_2 = 1000 \). (a) Combined 100 rpd plots of \( Y_1 \) (left-shifted curve) and \( Y_2 \) (right-shifted curve) vs. \( t \) for \( 0 \leq t < 10 \). (b) A 100 rpd plot of \( Y_1 \) vs. \( t \) for \( 0 \leq t < 30 \). (c) A 100 rpd plot of \( Y_2 \) vs. \( Y_1 \) for \( 0 \leq t < 30 \).
Henceforth, we shall specify the number of reactions that occur between each dot that appears on a simulation output graph by the notation
\[ \text{rpd} = \text{reactions per dot} \]  

Thus, Figure 3 is a 1 rpd plot, while Figure 4 is a 5 rpd plot. In Figure 5 we show a 10 rpd plot of a simulation run of reaction 22 for
\[ c = 0.5, \quad X_0 = 10000 \]  

(28)

Figures 3–5 illustrate two important, but certainly not unexpected, points: First, as the initial number of X molecules is increased, the temporal behavior of the system assumes more and more the appearance of the continuous, deterministic process described by the reaction-rate equation (23). Second, regardless of the initial number of X molecules, the temporal behavior as calculated by the stochastic simulation algorithm is entirely consistent with the predictions of the master equation (24). Of course, we are not attempting to prove here that the stochastic simulation algorithm provides a correct description of the time evolution of a chemically reacting system, for this has already been established in an a priori manner in sections II and III; rather, our purpose here is simply to demonstrate that this computational method is capable of being implemented. Figures 3–5 offer rather convincing proof of the feasibility of the stochastic simulation algorithm for the simple reaction 22. In this connection, we might note that the total cost of simulating and plotting the run shown in Figure 5 was about $5, the cost being split roughly equally between the simulating and plotting programs.

We turn next to a somewhat more complicated set of reactions:

\[ \bar{X} + Y \xrightarrow{c_1} 2Y \]  

(29a)

\[ 2Y \xrightarrow{c_2} Z \]  

(29b)

Here, the bar over X signifies that the molecular population level of this species is assumed to remain constant, either because the system is open to a large reservoir of X, or else because X is initially present in such great abundance that its depletion by reaction 29a will be negligible over any reasonable period of time.

The set of reactions (29) is of some interest because Malek-Mansour and Nicolis recently claimed that it provides a refutation of the basic stochastic hypothesis (4):

They proved that the master equation for reactions 29 is such that \[ \frac{\partial P(Y; t)}{\partial t} = 0 \] if and only if \[ Y = 0 \]; yet, the reaction-rate equation for (29)
\[ \frac{dY}{dt} = c_1 XY - 2(c_2/2)Y^2 \]  

(30)

\[ Y, = 0 \]  

(31a)

and
\[ Y^* = c_1 X/c_2 \]  

(31b)

and it can be shown that the first of these is mathematically unstable while the second is mathematically stable. This lead Malek-Mansour and Nicolis to conclude that the stochastic formalism upon which the master equation is based “seems to have destroyed the stable solution [31b] of the macroscopic equation [30] and preserved, instead, the physically unacceptable trivial (and unstable) solution [31a]: Even if the system starts initially with a great number of [Y] molecules, it will be driven eventually to extinction”.

Since the master equation and the stochastic simulation...
algorithm are logically equivalent to each other, we can test this conclusion simply by making a few simulation runs of reactions 29. For this, we need only apply the procedure outlined in Figure 2 with $M = 2, N = 1, X_1 = Y, h_1 = XY$, and $h_2 = Y(Y - 1)/2$. In Figure 6 we show a 10 rpd plot of two such runs with $c_1X = 5, c_2 = 0.005$ (Figure 6) for the initial conditions $Y = 10$ and $Y = 3000$. Contrary to what we might have expected from the result of Malek-Mansour and Nicolis, we see that both of these initial conditions lead to a situation in which the Y molecule population level fluctuates in a random but apparently stable manner over values in the vicinity of $c_1X/c_2 = 1000$, in reasonable agreement with the reaction-rate equation prediction (31b). In Figure 7 we show a 40 rpd plot of two more simulation runs of (29), these with $c_1X = 5, c_2 = 0.00125$ (Figure 7) for the two initial conditions $Y = 40$ and $Y = 12000$. Again, both initial conditions lead to stable random fluctuations over values in the vicinity of $c_1X/c_2 = 4000$. The only essential difference between the plots in Figures 6 and 7 is that the relative fluctuations in Figure 7 are about half as large as those in Figure 6: this is due to the fact that the molecular population levels in Figure 7 are four times larger than those in Figure 6.

The fact that the master equation for reactions 29 predicts a stable stationary state at $Y = 0$ can be easily understood directly from (29): clearly, if it should ever happen that the Y molecule population level ever drops to exactly zero, then that condition will persist indefinitely. However, it is very misleading to say that, if we start out with $Y > 0$, the system will be “driven” to $Y = 0$; for, as Figures 6 and 7 show quite convincingly, in such cases the Y molecule population level exhibits a definite affinity for values near $c_1X/c_2$. Of course, it is always possible that a chance, momentary, pairwise confluence of all the Y molecules could result in their complete annihilation via reaction 29b; and because this is possible, it will eventually happen if we just wait long enough ($t \to \infty$). However, this contingency is not especially significant from a macroscopic point of view; indeed, an examination of the trajectories in Figures 6 and 7 for $t > 2$ shows that, even for these relatively small numbers of molecules, one would have to wait an incredibly long time before observing a random fluctuation large enough to carry the system from the vicinity of $Y = c_1X/c_2$ down to $Y = 0$. More to the point here, though, is the fact that this contingency does not imply that the “transition probabilities [used in the stochastic approach] are not correctly written”, as was concluded by Malek-Mansour and Nicolis.

Shortly after the simulation runs in Figures 6 and 7 had been performed, the reader received a preprint of an article by Görtz and Walls, in which essentially these same conclusions were arrived at using an ingenious analytical technique based on the master equation. The reader is invited to consult their paper for further insights into this matter. Suffice it here to say that the objections to the stochastic formulation of chemical kinetics which Malek-Mansour and Nicolis have raised on the basis of their analysis of reactions 29 are now seen to be groundless. The fact that our numerical simulation algorithm was helpful in clarifying this essentially theoretical matter obviously speaks well for the potential usefulness of the algorithm.

In addition to providing information on if and how a steady state is approached, the stochastic simulation algorithm can also be used to estimate various statistical properties of the steady state. For example, returning to the situation illustrated in Figure 6 [reaction 29 with $c_1$ and $c_2$ as given in (32)], it is important to know just how well the population mean in the steady state is approximated by the reaction-rate equation estimate $c_1X/c_2 = 1000$; and it is also important to know how large the natural random fluctuations will be about that steady-state mean. Essentially what we are asking for here are the asymptotic values of the first moment $Y^{(1)}(t)$ and variance $\Delta^2(t)$ of the solution to the master equation for reactions 29. To obtain estimates of these values, we proceeded as follows: Starting with $1000 \ Y$ molecules at $t = 0$, we made a stochastic simulation run to $t = 0.3$, at which time we recorded the number of $\ Y$ molecules present; to simulate this length of system time required roughly 2000 reactions, and this was judged sufficient to make the system “forget” the initial number of $\ Y$ molecules. Repeating this process 4000 times, each time using different random numbers from URN (see Figure 2), we then estimated the mean and variance of the $\ Y$ values found at $t = 0.3$ by a straightforward averaging procedure. The results of these calculations were

$$Y^{(1)} = 999.9 \pm 1.2$$

where the ± uncertainties represent 95% confidence limits. The deterministic estimate of 1000 for the steady-state mean $Y^{(1)}$ is obviously well within our uncertainty limits. As for the steady-state variance $\Delta^2$, a subsequent analytical calculation furnished by Walls, using the computational procedure of Görtz and Walls, gave the result $\Delta^2 = (3/2)Y^{(1)}$ for reaction 22; in the present case this would imply $\Delta^2 = 1500$, which is obviously quite consistent with our result in (34).

The fact that the steady-state mean and variance are not equal to each other for reactions 29 is related to the fact that these reactions do not operate under conditions of chemical equilibrium (the reactions are irreversible, and they are also “driven” by the effective influx of X molecules). Another simple set of reactions which does not operate under conditions of chemical equilibrium is the set

$$X \xrightarrow{c_1} Y$$

$2Y \xrightarrow{c_2} Z$ (35b)

It has been shown by Nicolis, Mazo, and Görtz and Walls, using three different analytical techniques based on the master equation, that reactions 35 will have $\Delta^2 = (3/4)Y^{(1)}$. The significance of this result has been commented on by several authors. Taking $c_1X = 10^4$, $c_2 = 1$ (36)
the use of the corresponding reaction-rate equations

Several years later, Volterra independently investigated

possessed some remarkable dynamical properties.21

dYz/dt = c1Y1Y2 - c2Y1Yz - c3Yz 

(38a)

(38b)

(38c)

The computer programs used to obtain the statistical

estimates in (34) and (37) are easy to write, especially since they require no periodic write-outs for subsequent plotting. However, in performing this type of calculation, care must be taken to make each run long enough to wipe out effects due to the choice of the initial state. Moreover, as is typical of Monte Carlo estimates of averages, if one wishes to reduce the uncertainties by 1/f, one must make f2 times as many runs. These two features can conspire to make such calculations rather time consuming on a computer, and hence rather expensive; for example, it cost about $160 to obtain the figures quoted in (37). The chief advantage of the method is that it is universally applicable, and does not require complicated or specialized analytical techniques. The method is therefore best reserved for cases in which an analytical calculation either cannot be made at all, or else is of questionable validity because of various approximations employed.

IVB. The Lotka Reactions. In 1920, Lotka observed that the set of coupled, autocatalytic reactions

which yields a deterministic estimate for Y(1) of (c1X/c2)1/2 = 1000, we applied our stochastic simulation procedure to these reactions in essentially the same way that we did in obtaining the estimates in (34). The results were

Y(1) = 1000.7 ± 0.9 and Δ2 = 762 ± 35

(37)

where again the ± uncertainties represent 95% confidence limits. Our result for Δ2 is obviously consistent with the theoretically predicted value of 750.

Y1 + Y2 → Z

possessed some remarkable dynamical properties.21

Several years later, Volterra independently investigated the use of the corresponding reaction-rate equations

\[
\frac{dY_1}{dt} = c_1XY_1 - c_2Y_1Y_2 \\
\frac{dY_2}{dt} = c_2Y_1Y_2 - c_3Y_2
\]

(39a)

(39b)

to mathematically model a simple predator-prey ecosystem.22 Although the predator-prey interpretation of the Lotka reactions is a bit crude and cannot be pushed too far, it is helpful for visualizing and understanding the dynamics of these reactions. Thus, reaction 38b describes how a certain predator species Y2 reproduces by feeding on a certain prey species Y1; reaction 38a describes how Y1 reproduces by feeding on a certain foodstuff X, which is assumed here to be only insignificantly depleted thereby; and the isomerization 38c describes the eventual demise of Y2 through natural causes.

A steady state system would be characterized by the condition

dY1/dt = dY2/dt = 0

and it is easy to show from (39) that this condition is satisfied when

Y1 = Y_1s = c_2/c_1 and Y_2 = Y_2s = c_1X/c_2

(40)

That is, if we start out at t = 0 with Y1 = Y_1s and Y_2 = Y_2s, then the deterministic reaction-rate equations predict that this situation will persist indefinitely. However, as we shall now see, the physically more realistic stochastic approach tells quite a different story.

A stochastic simulation computer program for the Lotka reactions was written according to the scheme in Figure 2 (putting M = 3, N = 2, X_1 = Y_1, X_2 = Y_2, h_1 = XY_1, h_2 = Y_1Y_2, and h_3 = Y_2). Figures 8a, 8b, and 8c show three plots of one simulation run of this program with c_1X = 10, c_2 = 0.01, c_3 = 10 (Figures 8, 9)

(41)

for the initial condition Y_1 = Y_1s = 1000 and Y_2 = Y_2s = 1000. Figure 8a shows the Y_1 and Y_2 population levels plotted against time, at 100 rpd, over the interval 0 ≤ t ≤ 10; the leading (left-shifted) curve is the Y_1 (prey) population, and the lagging (right-shifted) curve is the Y_2 (predator) population. Instead of remaining constant at the 1000 level, as the reaction-rate equations (39) predict, the Y_1 and Y_2 populations are seen to develop a pronounced oscillatory behavior. These oscillations are ev-
identically characterized by a fairly stable frequency and phase difference, but a markedly unstable amplitude. Figure 8b is a 100 rpd plot of $Y_1$ only over the interval $0 \leq t \leq 30$, illustrating more completely the stability of the frequency and the instability of the amplitude. And Figure 8c is a 100 rpd plot over this same time interval showing the counterclockwise "orbits" traced out in the $Y_1$-$Y_2$ plane. The simulation run from which the plots in Figures 8a-c were constructed contained a total of $1 \times 10^5$ reactions. The cost of compiling and executing the simulation program for this run on the Univac 1110 computer was about $40, and the subsequent plotting programs averaged about $9 per graph.

Figures 9a, 9b, and 9c are 100 rpd plots of a second Lotka simulation run; this run was identical with the first except that a different chain of random numbers was used (i.e., the "starter number" for the unit-interval uniform random number generator was changed). A comparison of Figures 8a and 9a shows that, although the frequencies and the phase-differences are essentially the same in both runs, the amplitude variations are quite different; furthermore, at any particular instant (e.g., $t = 10$) the $Y_1$ and $Y_2$ values in Figure 9a are usually quite different from the $Y_1$ and $Y_2$ values in Figure 8a. Although it might seem from these two figures that the vagaries in the amplitude are less pronounced in the second run than in the first run, a comparison of Figures 8b and 9b shows that this is true only temporarily: between $t = 12$ and $t = 22$ in the second run, the population oscillations become quite extreme, ranging over two full orders of magnitude. The $Y_1$-$Y_2$ phase relationship in these large amplitude oscillations is illustrated in the 100 rpd plot in Figure 6c; the quasi-concentric phase plots as in Figures 8c and 9c, but he did subject his data to a detailed time-series analysis to determine the frequency components of the oscillations. As mentioned in section IIID, our stochastic simulation method represents an improvement over Nakaniishi's method in terms of both rigor and ease of application. Also, our results bring out somewhat more clearly than did Nakaniishi's results the relative ease with which the Lotka system can progress from the stationary state $(Y_1, Y_2)$ to the absorbing states $(0, 0)$ or $(\infty, 0)$.

Figure 10 is a 50 rpd plot of the Lotka reactions showing what happens when we "close" the system and let the $X$ population be depleted by reaction 38a in the normal way. The modification required in the simulation program to effect this depletion is trivial: in the section of the program that handles the predator population, we simply add an instruction to decrement the $X$ population by 1 after an $R_1$ reaction has occurred, we simply add an instruction to decrement the $X$ population by 1. In the run plotted in Figure 10 we used essentially the same parameters as in the previous Lotka runs, but we have allowed $X$ to decrease from an initial value of $10^5$:

$$c_1 = 0.0001, c_2 = 0.01, c_3 = 10$$

(Figures 10, 11)

The heavy appearance of the $X$ population trajectory is due to a "photographic burn" in the plotting hardware caused by the severe overlapping of successively plotted dots. The curve that oscillates about the 1000 level until
that the random number starter was changed. Again we see the extreme variability in the Lotka system: the vanishing of the Y2's occurs at t ≈ 28 in Figure 11, as compared with \( t \approx 19 \) in Figure 10; also, the final number of Y1's in Figure 11 turns out to be 7232, as compared with 16449 in Figure 10.

The behavior illustrated in Figures 10 and 11 suggests that a number of modifications could be made in the Lotka reactions (38) to make them a more realistic model of naturally occurring predator–prey systems. As a simple illustration, suppose we add a fourth reaction to (38a–c)

\[ Y_1 \rightarrow Z \]  

which allows the preys to die of natural causes in the same way as the predators. Taking

\[ c_1 = 0.0002, \quad c_2 = 0.01, \quad c_3 = c_4 = 10 \]  

(Figures 12, 13)

and starting with \( X = 10^5 \) and \( Y_1 = Y_2 = 10^4 \), two stochastic simulation runs were made. In one of these runs \( X \) was held constant, and in the other \( X \) was allowed to be depleted by reaction 38a. The results of the constant \( X \) run are plotted at 200 rpd in Figures 12a and 12b. These two plots are obviously very similar to the plots in Figures 8 and 9 (except that this modified run contains about half again as many reactions). Indeed, the reaction-rate equations for reactions 38a–d predict that, if \( X \) is held constant at \( 10^5 \) and \( c_1, \ldots, c_4 \) have the values in (43), then we will have \( \frac{dY_1}{dt} = \frac{dY_2}{dt} = 0 \) for \( Y_1 = Y_2 = 1000 \), just as we had in the runs in Figures 8 and 9. In Figure 13 we show a 50 rpd plot of the other simulation run, in which \( X \) is allowed to be depleted. As in Figures 10 and 11, the heavy curve is the \( X \) trajectory, the upper oscillating curve is the \( Y_1 \) trajectory, and the lower oscillating curve is the \( Y_2 \) trajectory. As expected, \( Y_1 \) as well as \( Y_2 \) becomes extinct in this case. However, it is intriguing to note that \( Y_2 \) becomes extinct considerably before \( Y_1 \) does, even though the natural life expectancy of a \( Y_2 \) is the same here as the natural life expectancy of a \( Y_1 \) (i.e., \( 1/c_4 = 1/c_3 \)). It is also interesting to note that over 40% of the initial amount of \( X \) remains after both \( Y_1 \) and \( Y_2 \) have become extinct.

It is obviously possible to modify the Lotka reactions in many other ways to obtain models of various other predator–prey systems. Suffice it here to say that, so long as these modifications can be expressed as additional chemical reactions and/or additional chemical species, they can be easily incorporated into our stochastic simulation computer program in a completely exact manner.

IVC. The Brusselator. The neutrally stable character of the oscillations in the Lotka model, as illustrated by the phase trajectory plots in Figures 8c and 9c, leads us to wonder if there exists a model set of coupled chemical reactions with “positively stable” oscillations. More precisely, is there a model chemically reacting system with two intermediate species \( Y_1 \) and \( Y_2 \) which has the property that, no matter what its initial state, the system will, as we had in the runs in Figures 8 and 9 (except that this modified run contains about half again as many reactions).

\[ X \leftarrow c_1 \rightarrow Y_1 \]  
\[ X + Y_1 \leftarrow c_2 \rightarrow Y_2 + Z \]  
\[ 2Y_1 + Y_2 \leftarrow c_4 \rightarrow 3Y_1 \]  
\[ Y_1 \leftarrow c_4 \rightarrow Z_2 \]  

\( c_1 = 0.0002, \quad c_2 = 0.01, \quad c_3 = c_4 = 10 \)  

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(Figures 12, 13)

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\[ 2Y_1 + Y_2 \leftarrow c_4 \rightarrow 3Y_1 \]  
\[ Y_1 \leftarrow c_4 \rightarrow Z_2 \]  

\( c_1 = 0.0002, \quad c_2 = 0.01, \quad c_3 = c_4 = 10 \)  

(Figures 12, 13)
Stochastic Simulation of Coupled Chemical Reactions

Figure 15. A 50 rpd plot of $Y_2$ vs. $Y_1$ for four stochastic simulation runs of the Brusselator (44), each run having the same reaction parameters as in Figure 14 but different initial values of $Y_1$ and $Y_2$.

Figure 16. Showing what would have happened in the run of Figure 14 if the system had been closed to species $X_1$. The exponentially decaying curve in (a) and (b) is $X_1$, and the oscillating $Y_1$ and $Y_2$ curve in (b) are both plotted at 100 rpd.

Figure 17. Showing what would have happened in the run of Figure 14 if the system had been closed to species $X_2$. The heavy, smooth curve is $Y_2$, and the horizontal jagged curve is $Y_1$, both plotted at 100 rpd.


This set of reactions has been dubbed the “Brusselator” by Tyson, and has been the object of extensive study by a number of workers. The deterministic reaction-rate equations for the Brusselator are evidently

\[ \begin{align*}
    \frac{dY_1}{dt} &= c_1 X_1 - c_2 X_2 Y_1 + (c_3/2) Y_1^2 Y_2 - c_4 Y_1 \\
    \frac{dY_2}{dt} &= c_2 X_2 Y_1 - (c_3/2) Y_1^2 Y_2
\end{align*} \]

(45a)  (45b)

and it is easy to show from these that $\frac{dY_1}{dt} = \frac{dY_2}{dt} = 0$ when $Y_1 = Y_{1s}$ and $Y_2 = Y_{2s}$, where

\[ Y_{1s} = c_1 X_1/c_4 \]

and

\[ Y_{2s} = 2c_2 X_2 c_4/c_1 X_1 c_3 \]

(46a)  (46b)

Following the prescription of Figure 2, a stochastic simulation computer program was written for the Brusselator (putting $M = 4$, $N = 2$, $X_1 = Y_1$, $X_2 = Y_2$, $h_1 = X_1 Y_2$, $h_2 = X_2 Y_1$, $h_3 = Y_3 Y_1(Y_1 - 1)/2$, and $h_4 = Y_1$). With $c_1 X_1 = 5000$, $c_2 X_2 = 50$, $c_3 = 0.00005$, $c_4 = 5$, the simulation run was made with $Y_1$ and $Y_2$ initially set to the values $Y_{1s} = 1000$ and $Y_{2s} = 2000$. Figures 14a and 14b are 100 rpd plots of $Y_1$ vs. $t$ and $Y_2$ vs. $t$, respectively, over the time interval $0 \leq t \leq 14$. Clearly, $Y_1$ and $Y_2$ do not remain at the stationary values $Y_{1s}$ and $Y_{2s}$, but commence almost immediately to oscillate vigorously in an apparently regular and stable way. The sharp rises in $Y_1$ in Figure 14a coincide with the sharp falls in $Y_2$ in Figure 14b so closely that they appear to be single vertical lines when the two figures are superposed. It is worth noting that these sudden, drastic changes in the molecular population levels pose no special problems to our stochastic simulation procedure. Figure 14c is a 50 rpd plot of $Y_1$ vs. $Y_2$ over the slightly longer time interval $0 < t < 18$; the cycle direction here is clockwise. The data in Figures 14a-c are from a $2 \times 10^6$ reaction simulation run, which took the Univac 1110 computer about 7 min to perform, at a cost of about $80.

Although Figure 14c shows that the phase orbits in the Brusselator are indeed much more stable than the phase orbits in the Lotka model [cf. Figures 8c and 9c], Figure 14c also shows a rather peculiar property of the Brusselator oscillations which, so far as the author is aware, has never been noticed before. This is the apparent inability of the system to consistently retrace its previous path on the diagonal leg of the limit cycle. The variability in these diagonal traces is directly related to the erratic variations in the heights of the peaks in Figures 14a and 14b. Although obviously caused by random fluctuations at the microscopic level, the effect itself is clearly more than microscopic: the relative variations in the peak heights in Figures 14a and 14b are on the order of 12%, which is a good order of magnitude larger than one would expect on the basis of the usual “square root” argument. The time intervals between successive oscillations, as determined by the distances between the successive vertical rise lines in Figure 14a, shows similar variations. Relatively crude ruler measurements on the graphs themselves indicate that the peak height achieved on any vertical rise in the $Y_1$ plot in
Figure 18. Results of a stochastic simulation run of the Brusselator with $c, x_1 = 5000, y_4 = 1000, y_5 = 2000,$ and $\alpha = 0.1,$ all plotted at 100 rpd: (a) $y_1$ vs. $t;$ (b) $y_2$ vs. $t;$ (c) $y_2$ vs. $y_1.$ Figure 14a is roughly proportional to the time elapsed since the previous rise. These observations would seem to warrant further investigations, although we shall not attempt any here.

The Brusselator phase trajectory in Figure 14c seems to have a different “character” along each of the three legs of the limit cycle. The peculiar character of the trajectory along the diagonal leg has already been commented on. Along the vertical leg the trajectory is fairly sharp and well-defined, subject to rather ordinary looking fluctuations. However, the trajectory along the horizontal leg has a thicker, labored look, with fluctuations that appear to be more “diagonal” than “up and down”. Indeed, it almost seems that the $y_1y_2$ plane is scored with a series of closely spaced, parallel diagonal grooves. Starting at the right end of the horizontal trajectory, the system phase point seems to struggle along against these grooves until it reaches the neighborhood of the point $(y_{1u}, y_{2u}).$ From there it seems to have a relatively smooth trip up the vertical leg, until it finally “drops” into one of the horizontal grooves. Precisely which groove the system phase point drops into is a matter of chance, but once lodged in one of these grooves it seems content to remain there for its quick trip down the final, diagonal leg of the orbit.

Figure 15 shows a 50 rpd plot of four short simulation runs, each having the same parameters (47) as in the previous run, but each starting out at different points in the $y_1y_2$ plane. As expected, no matter where we start out in this plane the system phase point eventually winds up on the clockwise limit cycle that we found in the previous run. However, the system seems to have very definite ideas as to just how it wants to approach the limit cycle: far from just spiraling in or out to the limit cycle as one might have naively expected, the system point seems again to be...
constrained by the imaginary diagonal grooves in the $Y_1Y_2$ plane mentioned above.

As we observed in our calculations on the Lotka model, it is very easy to modify our stochastic simulation program so as to "open" or "close" the system to any particular reactant species. As indicated in (44), the Brusselator is normally considered to be open to both $X_1$ and $X_2$; indeed, it is precisely this influx of $X_1$ and $X_2$ that drives the oscillator. In Figures 16a and 16b we show what happens when, given the parameter values in (47), we close the system to $X_1$, and allow $X_1$ to be depleted by reaction 44a from an initial value of $10^5$. In these two figures, the $X_1$ trajectory is plotted at 250 rpd while the $Y_1$ and $Y_2$ trajectories are each plotted at 100 rpd. In this case the system's behavior is not nearly so exotic: Starting out at $10^5$, $X_2$ quickly drops to zero—so rapidly in fact that an oscillating pattern for $Y_1$ and $Y_2$ does not have time to fully develop. $Y_2$, starting at 2000, decreases to zero, while $Y_1$ eventually settles back to its initial value 1000; this final steady state is easily understood from eq 46.

A stability analysis of the deterministic reaction-rate equations (45) by Lefever and Nicolis and also by Tyson predicts that the Brusselator should oscillate when the inequality

$$\frac{c_2X_2}{c_3} > \frac{c_1^2X_1^2}{c_4^2} + \frac{c_4}{c_3/2}$$

is satisfied, and presumably should remain at the stationary values $Y_2$ and $Y_1$ in (46) otherwise. To test this prediction, we first introduce a new parameter $\alpha$ defined...
by
\[ \alpha = 2c_4/c_2X_1 \]
(49)
From now on, instead of specifying the Brusselator reactions by the four parameters \( c_1X_1, c_2X_2, c_3, \) and \( c_4 \) as in (47), we shall use the four parameters, \( c_1X_1, Y_1, Y_2, \) and \( \alpha; \) this is permissible because, using (46) and (49)
\[ c_1X_1 = c_1X_1 \]
(50a)
\[ c_2X_2 = 2c_1X_1/\alpha Y_1 \]
(50b)
\[ c_3 = 4c_1X_1/\alpha Y_1^2 Y_2 \]
(50c)
\[ c_4 = c_1X_1/Y_1 \]
(50d)
Inserting (50) into the oscillation criterion (48), we find that this criterion assumes the somewhat simpler form
\[ \alpha < 1 \]
(51)
The significance of the new parametrization of the Brusselator reactions is that \( c_1X_1 \) controls the time scale of the reactions, \( Y_1 \) and \( Y_2 \) control the molecular population scales, and \( \alpha \) controls the stability of the reactions.

For \( Y_1 = 1000 \) and \( Y_2 = 2000 \), as in the runs plotted in Figures 14 and 15, the condition (51) for oscillatory instability in the Brusselator is simply
\[ \alpha < 2(1 - Y_1/Y_2) \]
(52)
Since (49) implies that \( \alpha = 0.2 \) for the parameter values in (47), then the oscillations observed in Figures 14 and 15 tend to confirm this criterion. To check this out more thoroughly, a series of six simulation runs was made with \( Y_1 = 1000 \) and \( Y_2 = 2000 \), and with \( \alpha \) assuming a succession of values from 1/10 to 10. The results of these runs are plotted in Figures 18–23:
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 0.1 \quad \text{(Figure 18)} \]
(53)
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 0.2 \quad \text{(Figure 19)} \]
(54)
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 0.5 \quad \text{(Figure 20)} \]
(55)
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 1.0 \quad \text{(Figure 21)} \]
(56)
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 2.0 \quad \text{(Figure 22)} \]
(57)
\[ c_1X_1 = 5000, \ Y_1 = 1000, \ Y_2 = 2000, \]
\[ \alpha = 10.0 \quad \text{(Figure 23)} \]
(58)
All these runs start with \( Y_1 = 1000 \) and \( Y_2 = 2000 \) at \( t = 0 \). The plots all have 100 rpd and use the same scale on the \( t \) axis; however, notice that Figures 18–20 use a different scale on the \( Y_1 \) and \( Y_2 \) axes than Figures 21–23. The \( \alpha = 0.1 \) plots in Figure 18 show the results of about \( 2 \times 10^9 \) reactions. The oscillations here are evidently quite violent. Notice in particular the incredible abruptness of the rises in \( Y_1 \) in Figure 18a and the concomitant falls in \( Y_2 \) in Figure 18b; e.g., near \( t = 10 \) the rise and fall lines are virtually parallel to the vertical right borders of the plots. Again we point out that this kind of rapid temporal population excursion poses no special problems for our stochastic simulation program. As we proceed from Figure 19 to 20, we observe that the amplitude of the oscillations decreases and their frequency increases, corresponding to the fact that \( \alpha \) is moving closer to the critical value at which we expect the oscillations to cease and stability to set in. Figure 21 shows the behavior exactly at the critical point \( \alpha = 1 \); the system evidently still oscillates, but the oscillations are ragged and uneven. Figure 22 shows the behavior for \( \alpha = 2 \), which is supposedly in the stable, nonoscillating regime; however, we see in Figure 22a that the \( Y_1 \) and \( Y_2 \) population levels, though holding

![Figure 22](image_url)

**Figure 22.** Same as Figure 21, except \( \alpha = 2.0 \).

![Figure 23](image_url)

**Figure 23.** Same as Figure 22, except \( \alpha = 10.0 \).
Stochastic Simulation of Coupled Chemical Reactions

Figure 24. Results of a $0.71 \times 10^6$ reaction stochastic simulation run of the Oregonator reactions (59), with $Y_1 = 500$, $Y_2 = 1000$, $Y_3 = 2000$, $v_1 = 2000$, and $r_2 = 50000$ [see (61)]. (a) Combined 100 rpd plots of $Y_1$ (low-peaking curve), $Y_2$ (middle-peaking curve), and $Y_3$ (high-peaking curve) vs. $t$ for $0 \leq t \leq 6$. (b) Same as (a), but for $1 \leq t < 3$. (c) A 250 rpd plot in $Y_1 Y_2 Y_3$ space for $0 \leq t \leq 6$. (d) Projection of the trajectory in (c) onto the $Y_1 Y_2$ plane. (e) Projection of the trajectory in (c) onto the $Y_2 Y_3$ plane. (f) Projection of the trajectory in (c) onto the $Y_1 Y_3$ plane.

fairly close to $Y_1$ and $Y_2$, respectively, still exhibit noticeable, anticorrelated excursions similar to those at the critical point (cf. Figure 21a). When we increase $\alpha$ to 10, in Figure 23, stability seems finally to be at hand.

IVD. The Oregonator. The Brusselator has been criticized as being physically unrealistic on the grounds that reaction 44c, requiring as it does a simultaneous collision among two $Y_1$ molecules and a $Y_2$ molecule, seems rather improbable from a kinetic theory point of view. However, Tyson and Light\textsuperscript{24} have shown that no set of chemical reactions with only two intermediate species ($Y_1$ and $Y_2$) can exhibit limit cycle oscillations unless it contains at least one reaction of order $\geq 3$. Field and Noyes\textsuperscript{26} have devised a model chemical oscillator which circumvents this difficulty by introducing a third intermediate species ($Y_3$); their model, which they have named the Oregonator, is\textsuperscript{20}

$$\begin{align*}
\bar{X}_1 + Y_1 &\rightarrow c_1 Y_1 \quad (59a) \\
Y_1 + Y_2 &\rightarrow C_2 Z_1 \quad (59b) \\
\bar{X}_1 + Y_1 &\rightarrow c_3 2Y_1 + Y_3 \quad (59c) \\
2Y_1 &\rightarrow C_4 Z_1 \quad (59d) \\
\bar{X}_1 + Y_2 &\rightarrow c_5 Y_1 \quad (59e)
\end{align*}$$

and evidently involves no reaction more complicated than

Figure 25. A 250 rpd plot in $Y_1 Y_2 Y_3$ space for four stochastic simulation runs of the Oregonator (59), each run having the same reaction parameters as in Figure 24 but different initial values of $Y_1$, $Y_2$, and $Y_3$. bimolecular.\textsuperscript{27} Field and Noyes originally devised the Oregonator as a highly idealized model of the Belousov-Zhabotinskii reactions, which involve the cerium ion catalyzed oscillatory bromate oxidation of malonic acid.\textsuperscript{28} A much more complete and realistic model of the Belousov-Zhabotinskii reactions was developed by Field, Köröš, and Noyes,\textsuperscript{29} and that model has been convincingly tested out by Edelson, Field, and Noyes\textsuperscript{30} through numerical integration of the corresponding reaction-rate equations. Our objective here is much less ambitious: we shall simply consider the Oregonator (59) as a chemical system in its own right, and show that its dynamical behavior for a given set of reaction parameters can be

begin by writing the deterministic reaction-rate equations numerically calculated with our stochastic simulation procedure.

To help guide us in assigning numerical values to the five reaction parameters $c_1 X_1$, $c_2$, $c_3 X_2$, $c_4$, and $c_5 X_3$, we begin by writing the deterministic reaction-rate equations for the Oregonator:

$$dY_1/dt = c_1 X_1 Y_2 - c_2 Y_1 Y_2 + c_3 X_2 Y_1 - 2(c_4/2)Y_1^2$$

$$dY_2/dt = -c_1 X_1 Y_2 - c_2 Y_1 Y_2 + c_3 X_2 Y_3$$

$$dY_3/dt = c_3 X_3 Y_1 - c_5 X_3 Y_3$$

If $Y_{1s}$, $Y_{2s}$, and $Y_{3s}$ are the values of $Y_1$, $Y_2$, and $Y_3$ for which $dY_1/dt = dY_2/dt = dY_3/dt = 0$ and if $\rho_1$ and $\rho_2$ are the deterministic reaction rates of the first two reactions (59a) and (59b) when these steady-state values obtain, i.e.

$$\rho_1 = c_1 X_1 Y_{3s}$$

$$\rho_2 = c_1 Y_{1s} Y_{2s}$$

then it is not difficult to show from (60) that

$$c_1 X_1 = \rho_{1s}/Y_{1s}$$

$$c_2 = \rho_{2s}/Y_{1s} Y_{2s}$$

$$c_3 X_2 = (\rho_{1s} + \rho_{2s})/Y_{1s}$$

$$c_4 = 2\rho_{1s}/Y_{1s}^2$$

$$c_5 X_3 = (\rho_{1s} + \rho_{2s})/Y_{2s}$$

Therefore, one way to specify values for the five reaction parameters $c_1 X_1$, $c_2$, $c_3 X_2$, $c_4$, and $c_5 X_3$ is to first specify values for the five auxiliary parameters $Y_{1s}$, $Y_{2s}$, $Y_{3s}$, $\rho_{1s}$, and $\rho_{2s}$. This is a convenient way to proceed because, by working with the parameters $Y_{1s}$, $Y_{2s}$, and $Y_{3s}$, we can more directly control the scales of the molecular population levels of the various species. Similar considerations motivated our reparametrization of the Brusselator reaction constants in (60).

Following the procedure outlined in Figure 2, a stochastic simulation computer program was written for the Oregonator (putting $M = 5$, $N = 3$, $X_1 = Y_1$, $X_2 = Y_2$, $X_3 = Y_3$, $h_1 = X_1 Y_2$, $h_2 = Y_1 Y_2$, $h_3 = X_2 Y_3$, $h_4 = Y_1(Y_{1s} - 1)/2$ and $h_5 = X_3 Y_3$). Taking

$$Y_{1s} = 500, Y_{2s} = 1000, Y_{3s} = 2000, \rho_{1s} = 2000, \rho_{2s} = 50000$$

the corresponding reaction parameters were calculated from (61), and a stochastic simulation run was made with $Y_1$, $Y_2$, and $Y_3$ initially set to the nominal steady-state values 500, 1000, and 2000, respectively. The results of this simulation run, which contained approximately 0.71 × 10^6 reactions, are shown in Figures 24a-f. Figure 24a is a 100 rpd plot of all three intermediate species vs. time. Just as we found in the Brusselator [cf. Figures 14a and 14b], the population levels quickly move away from the initial steady-state values and begin to oscillate vigorously in an apparently regular and stable manner. The low-peaking curve in Figure 24a is the $Y_1$ trajectory, the middle-peaking curve is the $Y_2$ trajectory, and the high-peaking curve is the $Y_3$ trajectory. To show the details of the cycling more clearly, the time interval $1 < t < 3$ is shown on a horizontally expanded scale in Figure 24b (which is also a 100 rpd plot). The cycling behavior shown here is quite consistent with the interpretation of Field and Noyes. When the $Y_2$ molecular population level is high and the $Y_1$ and $Y_3$ molecular population levels are low, reactions 59a and 59b dominate and act to slowly decrease the $Y_2$ population level; when the $Y_2$ population is finally reduced below a certain critical level (evidently in the neighborhood of 1000 according to Figure 24b), reaction 59c becomes momentarily dominant and produces a rapid increase in the $Y_1$ and $Y_3$ population levels; this increase in $Y_1$ and $Y_3$ in turn stimulates reaction 59d, which pulls $Y_2$ down again, and reaction 59e, which pulls $Y_2$ down and pushes $Y_3$ back up.

The cycle trajectories in $Y_1 Y_2 Y_3$ space are plotted at 250 rpd in Figure 24c. As we found in the Brusselator [cf. Figure 14c], we note that on some portions of the limit cycle the system phase point cannot consistently retrace its previous path. This seems to occur most prominently at the higher population levels, and is evidenced also in Figure 24a by the irregular variations in the peak heights of the $Y_1$, $Y_2$, and $Y_3$ curves. Although obviously due to random microscopic fluctuations, the overall effect is too large to be labeled microscopic. One possible explanation for this behavior might be that the cycling serves to transmit the relative fluctuations at the lower molecular population levels directly to the higher molecular population levels, where they are then much larger than would normally be expected. Figures 24d-f are 250 rpd plots of the projections of the phase trajectory onto the three phase planes; Figure 24d emphasizes the peculiar warped shape of the limit cycle orbit in $Y_1 Y_2 Y_3$ space.

Figure 25 is a 250 rpd plot of four short simulation runs which have the same parameters as in (62) but different starting points in $Y_1 Y_2 Y_3$ space; evidently, no matter where the starting point is taken, the system eventually winds up orbiting the same limit cycle path that was found in Figure 24c.

Figure 26 is a 100 rpd plot showing what happens when,
for the parameters in (62), the system is closed to species \( X_1 \), and the \( X_1 \) population level is allowed to be depleted by reaction 59a from an initial value of \( 10^4 \). The upper curve is the \( X_1 \) trajectory, and the low-peaking, middle-peaking, and high-peaking curves are as before the \( Y_1, Y_2, \) and \( Y_3 \) trajectories, respectively. The waves in the \( X_1 \) trajectory are caused by the \( Y_2 \) variations through reaction 59a. As \( X_1 \) decreases, the \( Y_1 \) and \( Y_3 \) oscillations occur farther apart in time, but the shape of their peaks evidently remains unaffected. The oscillation near \( t = 3.4 \) is the system’s last gasp; the action terminates at \( t = 11.5 \) with \( X_1 = Y_1 = Y_3 = 0 \) and \( Y_2 = 4470 \). Figure 27 is a 100 rpd plot showing what happens when we close the system to \( X_2 \) instead of \( X_1 \), and allow the \( X_2 \) population level to be depleted by reaction 59c from an initial value of \( 10^5 \). The waves in the \( X_2 \) trajectory are caused by the \( Y_1 \) variations through reaction 59c. In this case we see that the oscillations just slowly die out altogether. By time \( t = 6 \), about \( 0.25 \times 10^6 \) reactions have occurred; the action finally stops about \( 0.06 \times 10^6 \) reactions later, at \( t = 218 \), with \( Y_1 = Y_2 = Y_3 = 0 \) and \( Y_2 = 65 \).

Figures 28–30 show the effect of changing the Oregonator reaction parameters in such a way that the limit cycle oscillations are made to disappear. These figures show three separate simulation runs, all with \( Y_1 = 500, Y_2 = 1000, Y_3 = 2000 \) as in the previous runs, but with the other two parameters \( \rho_1 \) and \( \rho_2 \) set as follows:

\[
\begin{align*}
\rho_1 &= 2000, \quad \rho_2 = 20000 \quad (\text{Figure 28}) \quad (63) \\
\rho_1 &= 4000, \quad \rho_2 = 20000 \quad (\text{Figure 29}) \quad (64) \\
\rho_1 &= 5000, \quad \rho_2 = 50000 \quad (\text{Figure 30}) \quad (65)
\end{align*}
\]

Comparing these figures with their identically plotted counterparts in Figures 24a and 24c (which are for \( \rho_1 = 2000 \) and \( \rho_2 = 50000 \)) shows that, as the ratio \( \rho_1/\rho_2 \) is increased from 0.04 to 1.00, the regular oscillations dissolve into random microscopic fluctuations about the nominal steady-state levels.

Finally, to investigate the effects of moving somewhat closer to the high molecular population levels realized experimentally, we made a stochastic simulation run of the Oregonator in which all the population levels were uniformly increased by a factor of 4 over the run plotted in Figure 24. This was accomplished by choosing the reaction parameter values so that, in contrast to (62)

\[
\begin{align*}
Y_1 &= 2000, \quad Y_2 = 4000, \quad Y_3 = 8000, \quad \rho_1 = 2000, \quad \rho_2 = 50000 \quad (\text{Figure 31}) \quad (66)
\end{align*}
\]

Figure 31a is a 500 rpd plot showing the details of the cycling, and is to be compared with Figure 24b. Figure 31b is a 1000 rpd plot of the phase trajectory in \( Y_1, Y_2, Y_3 \) space, and is to be compared with Figure 24c. Evidently, the only significant difference between these plots is the fourfold increase in the molecular population levels, and an accompanying decrease in the relative size of the random, microscopic fluctuations. Especially noteworthy is the fact that the curious "spreading" of the limit cycle path in \( Y_1, Y_2, Y_3 \) space does not seem to have been appreciably reduced; certainly, this spreading is much greater than the microscopic fluctuations on the individual orbits. The fourfold increase in the molecular population levels in this run was attended by a fourfold increase in the numbers of reactions per cycle, and hence a fourfold increase in the execution time of the computer simulation program: Figure 31b shows the results of approximately \( 2.8 \times 10^9 \) reactions, which cost about $80 to simulate to the Univac 1110.

In view of the consistency between Figures 24b and 31a, and between Figures 24c and 31b, there would seem to be little point to increasing the molecular population levels any further. In this connection, it should be recognized...
that it is not necessarily desirable to scale the molecular population levels so high that all trajectories appear to be devoid of random fluctuations. Such fluctuations should be regarded as an intrinsically real aspect of the dynamical behavior of a chemically reacting system, not as some imperfection in our observational technique that ought to be eliminated. Indeed, part of the utility of our stochastic simulation procedure is that it affords us a unique opportunity to examine these fluctuations in a wide variety of controlled situations.

V. Concluding Remarks

This paper has described and illustrated the use of a novel method for numerically calculating the time evolution of a spatially homogeneous system of coupled chemical reactions. This "stochastic simulation algorithm" has been shown to be physically and mathematically well-grounded from a kinetic theory point of view, and rigorously equivalent to the spatially homogeneous master equation, yet surprisingly simple and straightforward to implement on a digital computer. The principle advantages and limitations of the algorithm have been summarized in section IID. Suffice it here to say that the specific applications considered in section IV demonstrate quite convincingly both the feasibility and the potential usefulness of this computational approach.

A comparison of the relative advantages of the stochastic simulation algorithm with those of the traditional method of numerically solving the deterministic reaction-rate equations is perhaps inevitable, and strong points can be scored on both sides. However, it would seem more fruitful to focus on the complementary aspects of these two computational methods instead of the competitive aspects. From this point of view, each method represents a tool, with specific advantages and limitations, which the chemical kineticist may employ at his discretion to elucidate the temporal behavior of a chemically reacting system.

As presented here, the stochastic simulation algorithm is applicable only to spatially homogeneous systems. Work toward extending the algorithm to accommodate molecular diffusion in spatially inhomogeneous systems is currently in progress, and will be reported on at a later date.

Acknowledgment. The numerical results reported in this paper were calculated on the NWC Central Computing Facility's Univac 1110 computer, and were plotted with their versatile DISSPLA/FR-80 computerized plotting system; the author is especially grateful to Hans Lindblom of the CCF for his helpful guidance in the use of the latter. The author acknowledges with thanks some fruitful discussions and correspondence with Dr. Dan Walls and Dr. Crispin Gardiner of the University of Waikato, and also some clarifying remarks by Professor N. G. van Kampen and Dr. G. Nicolis. Finally, the author takes special pleasure in expressing his gratitude to his wife Carol, whose generous advice in her capacity as a professional scientific programmer made the author's efforts on the computer much more productive and enjoyable than they would have been otherwise.

References and Notes

(2) In going from (2a) to (2b), and also from (3b) to (3c), it might appear that we are suggesting that an imminent collision is possible even between two molecules that are very far apart. However, this is not implied here. For a detailed discussion of this point, see D. T. Gillespie, J. Stat. Phys., 18, 311 (1977).
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(3) D. A. McQuarrie, J. Appl. Prob., 4, 413 (1967).

(4) This average is equivalent to the "ensemble average" that was denoted by angular brackets in eq {5a} and {6b}.

(5) It is considered implicit in (4) that the probability for more than one reaction to occur in d{t} is of order {1/dt}, and is therefore effectively zero. Thus, between times t and t + dt, either no reactions will occur, or else some one of the M{R} reactions will occur.

(6) The reaction-rate equations (eq {1}) have only one independent variable, namely, the continuous variable t; however, the master equation (eq {13}) has N = 1 independent variables, namely the N discrete variables X{1}, ..., X{N} and the one continuous variable t.

(7) For a somewhat more rigorous version of the foregoing arguments, see section III of ref 1.

(8) The art of writing reliable unit-interval uniform random number generators is quite involved, and is best left to the experts; see, for example, D. E. Knuth, "The Art of Computer Programming", Vol. 2, Addison-Wesley, Reading, Mass, 1969, and also G. Marsaglia and T. A. Bray, Commun. Assoc. Comput. Mach., 11, 757 (1968). It is important to note that these generators, even those written in Fortran, are usually machine dependent; a Fortran generator written for one kind of computer will seldom work properly on another kind of computer.


(12) The discrete character of our {r}-selection process is not a result of any approximation scheme, but merely a consequence of the fact that any bounded chemical system contains a finite number of molecules.


(15) D. F. Walle, private communication.


(20) A bar over any reactant species signifies that its molecular population is assumed to remain constant; the system is said to be "open" to that species.


(22) For a comprehensive review of Volterra's and some nonlinear models of interacting populations, see N. S. Goel, S. C. Maitra, and E. W. Montroll, Rev. Mod. Phys., 43, 231 (1971).


(27) Field and Noyes actually write the fifth reaction as Y{3} + Y{4} \rightarrow Y{5}, where f{A} is a "pychimotic factor" which is introduced in an attempt to obtain a better correspondence with the real Belousov-Zhabotinski reactions. In most cases, as here, f{A} is simply taken as unity. The "reversible Oregonator" has been discussed by R. J. Field, J. Chem. Phys., 83, 2266 (1980); the "closed Oregonator" has been discussed by R. M. Noyes, J. Chem. Phys., 64, 1266 (1976).


(31) From our point of view, eq {51} do not assume the validity of the deterministic reaction-rate equations (eq {60}); rather, they offer an alternate parametrization of the Oregonator reaction constants.

Discussion

W. C. GARDINER, JR. (University of Texas). (1) Your stochastic algorithm involves substantially more operations per "integration cycle" (one more call to URN and one to ALOG) than the procedure published by Bunker and co-workers (Combust. Flame, 1974). What does your procedure gain from this extra work? (2) What is the largest ratio of species concentrations that can reasonably be handled in a practical application of your routine?

D. T. GILLESPIE. (1) The relationship between my stochastic simulation algorithm and the procedure of Bunker et al. (ref 11) is discussed in section IIIID of this paper, and also on p 429 of ref 1. The main point is that my method is fully equivalent to the spatially homogeneous master equation, whereas the method of Bunker et al. is not. Therefore, a more appropriate question would be: "Under what conditions may one replace, in step 2 of Figure 2, the statement t = (1/\alpha) \ln (1/r{\alpha}) with the simpler statement t = (1/\alpha), without introducing sensible alterations in the computational results?" I think it should certainly be possible to make this replacement when one is calculating the mean and variance of a stable or metastable state, as described in connection with eq 34 and 37; in fact, for that type of calculation one could probably just dispense with t altogether. However, for calculating a system's transient behavior, I do not see how one could confidently answer this question except empirically on a case-by-case basis. I am sure that for many situations the shorter expression for t would prove to be adequate; but notice, for example, that it could not be used for the simple isomerization reaction 22 if one were interested in studying fluctuations. I have made no study of this matter in these initial calculations, essentially taking the position that the a priori assurance of being rigorously correct, which comes with using the longer expression for t, is worth the fractional increase in computer running time. (2) The runs plotted in Figures 16 and 18 are "reasonable", and show relative population ratios that span roughly four orders of magnitude. Of course, the fundamental limitation on this procedure is not the population ratios per se, but rather the total number of individual reaction events that must be simulated in order to cover the time span of interest. At typical charging rates on the Univac 1110, I found a rough rule of thumb to be $30 per 10^6 reactions. The runs plotted in Figures 16 and 18 contain between 1 and 2 \times 10^6 reactions each.

ROBERT J. GELINAS (Science Applications, Inc.). I must take exception to the remarks made subsequent to Gillespie's talk by Professor Shuler, and the other allowed questioner, disparaging Gillespie's method. Gillespie's general method has strong foundations in quantum kinetic theory and in corresponding Langevin analysis which were not discussed. His imaginative reaction accounting method is to conventional statistical approaches based on species concentrations, as second quantization is to first quantization in quantum theory. I cannot vouch for his algebra at this time, but his basic method is worthy of positive acknowledgment and encouragement. As with the originations of many new creative ideas, time is required to develop a venge against unwarranted criticism. It would certainly be a disservice to discourage further pursuit of Gillespie's work at this early time.